



Contents lists available at ScienceDirect

## Environmental Pollution

journal homepage: [www.elsevier.com/locate/envpol](http://www.elsevier.com/locate/envpol)

# Metal(loid) oxides and metal sulfides nanomaterials reduced heavy metals uptake in soil cultivated cucumber plants<sup>☆</sup>

Chun Song<sup>a,\*</sup>, Fang Ye<sup>a</sup>, Huiling Zhang<sup>b</sup>, Jie Hong<sup>c</sup>, Chenyu Hua<sup>b</sup>, Bin Wang<sup>b</sup>,  
Yanshan Chen<sup>b</sup>, Rong Ji<sup>b</sup>, Lijuan Zhao<sup>b</sup>

<sup>a</sup> Institute of Ecological and Environmental Sciences, Sichuan Agricultural University, Chengdu, 611130, China

<sup>b</sup> State Key Laboratory of Pollution Control and Resource Reuse, School of Environment, Nanjing University, Nanjing, 210046, China

<sup>c</sup> College of Environment, Zhejiang University of Technology, Hangzhou 310014, China

## ARTICLE INFO

## Article history:

Received 21 May 2019

Received in revised form

30 August 2019

Accepted 5 October 2019

Available online 7 October 2019

## Keywords:

Contaminated soil

Nanomaterials

Heavy metals

Mineral nutrients

Cucumber

## ABSTRACT

Agricultural soil is one of the main sink for both heavy metals and nanomaterials (NMs). Whether NMs can impact heavy metals uptake or bioaccumulation in plants is unknown. Here, cucumber plants were cultivated in a multi-heavy metals contaminated soil amended with four types of NMs (SiO<sub>2</sub>, TiO<sub>2</sub>, ZnS and MoS<sub>2</sub>) separately for four weeks. Physiological and biochemical parameters were determined to investigate the impact of NMs on plant growth. Inductively coupled plasma mass spectrometry was employed to determine the metal content in plants. Results showed that none of the tested NMs impacted plants biomass, but all the NMs showed different degrees of reduction in heavy metals bioaccumulation in plant roots, stems and leaves. However, four NMs showed different degrees of reduction in macro and micro nutrients uptake. MoS<sub>2</sub> decreased the bioaccumulation of heavy metals (As, Cd, Cr, Cu, Ni, Al, Ti and Pb) for 36.4–60.6% and nutrients (Mg, Fe, K, Si and Mn) for 40.1%–50.1% in roots. Exposure to MoS<sub>2</sub> NMs also significantly increased 23.4% of Si in leaves, 205.6% and 83.9% of Mo in roots and stems, respectively. In general, the results of this study showed promising potential for NMs to reduce uptake of heavy metals in crop plants, especially MoS<sub>2</sub> NMs. However, the negative impacts of perturbing nutrients uptake should be paid attention as well.

© 2019 Elsevier Ltd. All rights reserved.

## 1. Introduction

Due to the large amount of pollutants discharge from mining and manufacturing industries, soil contamination is becoming an important issue globally (Cachada et al., 2018). Unlike organic contaminants, metals or metalloids do not undergo microbial or chemical degradation and will be persist for a long time in soil (Bolan et al., 2014). Crops grown in heavy metal contaminated soil might produce unsafe food, posing threats to human health (Zhu et al., 2007). Using soil amendments to decrease the bioavailability or mobility of metal/metalloid is an important strategy to reduce metal accumulation in crops (González et al., 2013). A variety of materials have been used as immobilization amendment, such as phosphate compounds, liming materials, organic composts, metal oxides and biochars (Bolan et al., 2014; Yue et al., 2019).

Recently, some engineered nanomaterials (NMs) have been applied for environmental remediation due to their capacities for heavy metal absorption and high surface area (Lehman and Larsen, 2014). For example, the core (iron oxide NMs)-shell (mesoporous silica) structure enable the separation of the nanocomposites from water using permanent magnet (Chen et al., 2009). Magnetic mesoporous silica nanocomposites have been used for heavy metals (Cr and Cu) decontamination (Lin and Haynes, 2009; Chen et al., 2011). ZnS NMs was used for the removal of Pb<sup>2+</sup> and Hg<sup>2+</sup> from aqueous solutions (Pala and Brock, 2012). MoS<sub>2</sub> can adsorb heavy metal ions due to the strong Lewis acid and base soft-soft interactions (Wang and Mi, 2017). Silicon nanoparticles have been shown to reduce chromium (Cr) accumulation in *Pisum sativum* seedlings (Tripathi et al., 2015). TiO<sub>2</sub> NMs have been reported to significantly reduce heavy metals uptake in benthic organisms (*Corbicula fluminea*) (Fan et al., 2018). However, most of these studies were conducted in water body, only a few studies focused on contaminated soil remediation using NMs. It was recently reported that Fe<sub>3</sub>O<sub>4</sub> NMs reduced heavy metals (As, Pb, Zn, Cd and Cu)

<sup>☆</sup> This paper has been recommended for acceptance by Baoshan Xing.

\* Corresponding author.

E-mail address: [songchun@sicau.edu.cn](mailto:songchun@sicau.edu.cn) (C. Song).

uptake and alleviated their toxicity in plant seedling (Konate et al., 2017; Praveen et al., 2018).

On the other hand, engineered nanomaterials are regarded as emerging contaminants, which started to be produced in past two decades (Stark et al., 2015). The un-intended co-existence of heavy metals and nanoparticles in soil will be potentially increased due to the increasingly released NMs to agricultural soils through waste water irrigation and sewage sludge reuse. Therefore, to investigate the impact of NMs on crop plant performance is imperative. Most studies used uncontaminated soil to conduct the experiments, and little is known about the impact of NMs on heavy metal uptake and impact on plant growth in contaminated soil. Our hypothesis is that nanoparticles may reduce the uptake and bioaccumulation of heavy metals in plant, by binding or complexing with heavy metals, and thus promoting the plant growth.

In this study, we chose two commonly used adsorbents ( $\text{SiO}_2$  and  $\text{TiO}_2$  NMs) and two uncommonly used, but exhibiting great potential as adsorbents ( $\text{ZnS}$  and  $\text{MoS}_2$  NMs) to test their performance on heavy metals uptake in plant grown in previously contaminated soil. Another reason for selecting those NMs is due to their wide application in various industries and their potential release into environment is estimated to be increasing. Cucumber is an economically important vegetable crop. Meanwhile, cucumber (*Cucumis sativus*) have been reported to be sensitive to a wide range of contaminants (Gorsuch et al., 1991). Thus, cucumber plants were selected as model system to conduct the experiment. In this study, cucumber plants were allowed to grow in heavy metals contaminated soils collected from mining area for 4 weeks. Four types of NMs ( $\text{SiO}_2$ ,  $\text{TiO}_2$ ,  $\text{ZnS}$ ,  $\text{MoS}_2$ ) were amended in soil with a final dosing as 100 mg/kg. Physiological and biochemical response of cucumber plants to these NMs were measured. Meanwhile, the effect of NMs on heavy metals (As, Cd, Cr, Pb) and nutrient elements (Fe, Ca, K, Mg, Zn) uptake and bioaccumulation in cucumber plants were also determined. The approached data will be valuable for the risk assessment of heavy metals, NMs, or heavy metals/NMs co-exposure in environment.

## 2. Materials and methods

### 2.1. Nanomaterials and soil

$\text{SiO}_2$  and  $\text{TiO}_2$  NMs were purchased from Pantian Nano Inc. (Shanghai, China) and the original size is 20 and 5–10 nm, respectively.  $\text{ZnS}$  and  $\text{MoS}_2$  NMs were purchased from Gangtian Inc. (Shanghai, China) and Bohan Inc. (Shanghai, China) and the original size is 50 and 120 nm, respectively. The hydrodynamic size for  $\text{SiO}_2$ ,  $\text{TiO}_2$ ,  $\text{ZnS}$  and  $\text{MoS}_2$  are  $876 \pm 41.8$ ,  $801 \pm 11.4$ ,  $3825 \pm 492$  and  $1046 \pm 69.7$  nm, with the zeta ( $\zeta$ ) potential of  $25.0 \pm 0.666$ ,  $-17.5 \pm 0.476$ ,  $19.0 \pm 1.485$  and  $-29.0 \pm 0.592$  mV, measured via dynamic light scattering (Malvern Zetasizer Nano ZS-90), in nano pure water at pH 7 with 100 mg/L in nano pure water. The data of size and surface charge were summarized in Table 1.

The soil was collected from a field ( $32^\circ 09' 30.29'' \text{N}$ ,  $118^\circ 56' 56.95'' \text{E}$ ) in a mining area in Qixia Mountain (Nanjing, China), from the top 20 cm. The soil has been contaminated by a nearby lead-zinc mining factory, which was one of the largest lead

zinc deposits in East China and has been explored for more than 50 years. The collected soil was air-dried and ground to pass through a 2-mm sieve for the pot trial. ICP-MS analysis showed that the most abundance heavy metals in soil are As ( $65.2 \pm 3.0$  mg/kg), Pb ( $182 \pm 6.0$  mg/kg) and Cd ( $3.52 \pm 2.02$  mg/kg), respectively. Soil pH is  $6.46 \pm 0.093$ .

### 2.2. Exposure experiments

Cucumber (*Cucumis sativus*) seeds (Zhongnong 28 F1) were purchased from Hezhiyuan Seed Corp. (Shandong, China). The soil was spiked with different nanomaterials ( $\text{SiO}_2$ ,  $\text{TiO}_2$ ,  $\text{ZnS}$ ,  $\text{MoS}_2$ ) by mixing the NMs with a small portion of soil before blending the mixture with remaining soil homogeneously. Based on our previous studies and other studies used 500–1000 mg/kg NPs, which is a too high dose. Therefore, we were attempted to use a relatively low dose. The final dose of NMs in soil is 100 mg/kg. Cucumber seeds were planted in pots containing 100 g of soil. Each pot contained one seed. The plants were cultivated in a greenhouse for 28 days at  $25^\circ \text{C}$  during the day and  $20^\circ \text{C}$  at night. The daily light integral was  $180 \mu\text{mol m}^{-2} \cdot \text{s}^{-1}$  in the daytime. During the growth period, the plants were watered as needed and no additional fertilizers were applied.

### 2.3. Biomass and chlorophyll content analysis

At harvest, cucumber plants were thoroughly rinsed with tap water for 5 min followed by deionized water for 3 times. The fresh biomass of stem and leaf tissues was determined before oven-drying ( $70^\circ \text{C}$  for 72 h). Due to part of the roots was lost while washing, the root biomass was not calculated. Photosynthetic pigment analysis was performed to determine the levels of chlorophyll *a* and *b* and total carotenoids. The photosynthetic pigment contents were extracted following the protocol of Šesták et al. (1971). Briefly, 0.01 g of cucumber young leaves were mixed with 5 mL of 80% methanol for 12 h, and then the mixture was centrifuged for 10 min at 3000 rpm. A microplate Reader (Biotek Synergy H1, America) was used to measure the absorbance of chlorophyll *a* and *b* and carotenoids in the methanolic extracts at 666, 653, and 470 nm, respectively.

### 2.4. Total phenolics content

The total content of phenolic compounds were determined with the method of Singleton and Rossi (1964). Specifically, 50  $\mu\text{L}$  of the methanolic extract was diluted with 450  $\mu\text{L}$  of deionized (DI) water, and then 250  $\mu\text{L}$  of 2 M Folin–Ciocalteu reagent and 1.25 mL of 20 g/L  $\text{Na}_2\text{CO}_3$  were added to the mixture. After centrifugation, the absorbance of the supernatant was determined at 735 nm with UV spectra. Quality assurance and quality control include duplicated samples at 10%.

### 2.5. Lipid peroxidation

Lipid peroxidation in leaves was measured by the Thiobarbituric Acid Reactive Substances (TBARS) assay (Jambunathan, 2010). Malondialdehyde, which is the final product of fatty acid degradation, is an indicative of lipid peroxidation. Briefly, 0.2 g of fresh cucumber leaves were mixed with 4 mL of 0.1% trichloroacetic acid (TCA); the mixture was then centrifuged at 10,000 rpm for 15 min. A 1-ml aliquot of the supernatant was mixed with 2 mL of 20% TCA and 2 mL of 0.5% thiobarbituric acid (TBA); then, the mixture was heated in water bath at  $95^\circ \text{C}$  for 30 min. After cooling, the visible light absorbance was measured at 532 nm and 600 nm (Biotek

**Table 1**  
Size and  $\zeta$  Potential of nanomaterials (100 mg/L).

|                | Original Size (nm) | Hydrodynamic Size (nm) | Zeta Potential (mV) |
|----------------|--------------------|------------------------|---------------------|
| $\text{SiO}_2$ | 20                 | $876 \pm 41.8$         | $25 \pm 0.666$      |
| $\text{TiO}_2$ | 5–10               | $801 \pm 11.4$         | $-17.5 \pm 0.476$   |
| $\text{ZnS}$   | 50                 | $3825 \pm 491.8$       | $19.0 \pm 1.485$    |
| $\text{MoS}_2$ | 120                | $1046 \pm 69.7$        | $-29 \pm 0.592$     |

Synergy H1, America). Lipid peroxidation was expressed as  $\mu\text{mol}$  MDA equivalents  $1\text{ g}^{-1}$  of fresh weight.

## 2.6. ICP-MS analysis for element content in plant tissues

Root, stem and leaf for metal and metalloid analysis were dried at  $70\text{ }^{\circ}\text{C}$  for 72 h. A sample of approximately 0.02 g of dried tissue was microwave-digested (Milestone, Ethos Up, Germany) in a mixture of 8 mL of  $\text{H}_2\text{O}_2$  and 2 mL of  $\text{HNO}_3$  (4/1 v/v) at  $160\text{ }^{\circ}\text{C}$  for 40 min. The resulting digest was diluted to a final volume of 50 mL prior to analysis. The content of Si, Ti, Zn, Mo, macro- and micro-nutrients (K, Ca, Mg, Cu, Na and Mn) and heavy metals (As, Pb, Cr, Cd) were quantified by inductively coupled plasma mass spectrometry (ICP-MS) (NexION-300, PerkinElmer, USA) and inductively coupled plasma-optical emission spectrometer (ICP-OES) (Optima 8300, PerkinElmer, USA). Standard reference materials from National Institute of Standards and Technology 1547 and 1570a were used to validate the digestion and analytical method obtaining recoveries between 90% and 99%. The detection limit of Si, Ti, Zn, Mo, K, Ca, Na, Mg, Cu, Ni, Al, Mn, As, Pb, Cd, and Cr quantified by ICP-MS analysis were  $0.1\text{ }\mu\text{g L}^{-1}$ ,  $0.0008\text{ }\mu\text{g L}^{-1}$ ,  $0.1\text{ }\mu\text{g L}^{-1}$ ,  $0.001\text{ }\mu\text{g L}^{-1}$ ,  $1.0\text{ }\mu\text{g L}^{-1}$ ,  $1.9\text{ }\mu\text{g L}^{-1}$ ,  $0.6\text{ }\mu\text{g L}^{-1}$ ,  $0.1\text{ }\mu\text{g L}^{-1}$ ,  $0.009\text{ }\mu\text{g L}^{-1}$ ,  $0.02\text{ }\mu\text{g L}^{-1}$ ,  $0.03\text{ }\mu\text{g L}^{-1}$ ,  $0.03\text{ }\mu\text{g L}^{-1}$ ,  $0.03\text{ }\mu\text{g L}^{-1}$ ,  $0.0004\text{ }\mu\text{g L}^{-1}$ ,  $0.01\text{ }\mu\text{g L}^{-1}$  and  $0.01\text{ }\mu\text{g L}^{-1}$ , respectively.

## 2.7. Soil pH determination

The mixture of 3 g soil and 30 mL nano pure water was shaken for 2 h at  $20\text{ }^{\circ}\text{C}$  with 200 rpm. Then the pH of the suspension was measured using a pH meter (Mettler-Toledo, Switzerland).

## 3. Results and discussion

### 3.1. Impact of NMs on photosynthetic pigments content and biomass

Results showed that  $\text{SiO}_2$  and  $\text{TiO}_2$  NMs had no impact on photosynthetic pigment contents in cucumber leaves (Fig. 1). The previous studies from our group also found that neither  $\text{SiO}_2$  nor  $\text{TiO}_2$  NMs impacted photosynthetic pigment contents and growth

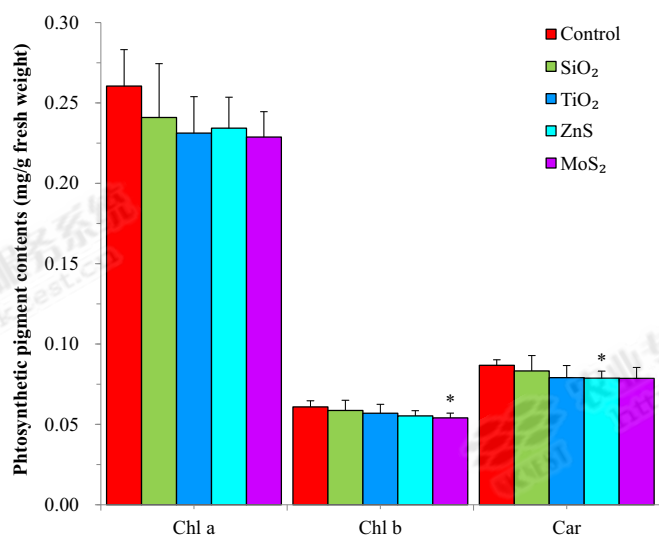


Fig. 1. Impact of Nanomaterials ( $\text{SiO}_2$ ,  $\text{TiO}_2$ , ZnS and  $\text{MoS}_2$ ) on photosynthetic pigment contents in cucumber plants. All data are average  $\pm$  SD of four replicate. \* represent statistical differences at  $p \leq 0.05$  compared with control, respectively.

of maize plants grown in another agricultural soil (Mollisol) dosing at  $100\text{ mg/kg}$  (Zhao et al., 2019). Song et al. (2013) also found that  $\text{TiO}_2$  NMs did not exhibit negative impact on chlorophyll content and growth of three plant species in hydroponic condition. Although not statically significant, exposure to  $\text{TiO}_2$  and  $\text{MoS}_2$  NMs distinctly decreased carotenoids by 8.84% ( $p = 0.082$ ) and 9.37% ( $p = 0.077$ ) compared to control, respectively. In contrast, exposure to ZnS NMs significantly decreased carotenoids by 9.18% ( $p \leq 0.05$ ) compared to control (Fig. 1). In addition, chlorophyll *b* content was also markedly decreased by 9.2% ( $p = 0.064$ ) upon ZnS exposure. These results indicate that ZnS NMs have negative impact on photosynthetic pigment biosynthesis of cucumber plants. Similarly, exposure to  $\text{MoS}_2$  NMs significantly decreased chlorophyll *b* content by 11.33% ( $p \leq 0.05$ ) compared to control (Fig. 1). Taken together, both metal sulfide NMs (ZnS and  $\text{MoS}_2$ ) inhibited photosynthetic pigment contents. The negative impact of NMs on photosynthetic pigment contents is in the order of  $\text{MoS}_2 \approx \text{ZnS} > \text{SiO}_2 \approx \text{TiO}_2$ . Since the leaf is not the organ which were directly exposure to NMs, the reason for the decreased photosynthetic pigments are probably because of the binding/complexing of NMs with nutrient elements (Mg and Fe) in the rhizosphere, which reduced the uptake of photosynthetic related elements by leaves and inhibited chlorophyll content. Further investigate is needed to verify this hypothesis. The changes of photosynthetic pigment content may influence the biomass accumulation. However, none of the tested NMs ( $\text{SiO}_2$ ,  $\text{TiO}_2$ , ZnS and  $\text{MoS}_2$ ) showed impacts on cucumber plants biomass (stem and leaf) during 4 weeks exposure (Fig. 2A), even in plants exposed to ZnS and  $\text{MoS}_2$  NMs in which photosynthetic pigment contents were negatively impacted. Also, no visible toxicity symptoms were observed during the whole period of cultivation (Fig. 2B). Taken together, all the tested NMs generally had no impact on plant growth.

### 3.2. Impact of NMs on leaf lipid peroxidation and total phenolics

Nanoparticle can either directly interact with root, or the released ions from NPs can be taken up by root and induce metabolic changes. To test whether NMs or ions induce lipid peroxidation, malondialdehyde (MDA) content, which is the reaction product of membrane lipid peroxidation, were determined. Results showed that MDA content was unchanged by all types of NMs (Fig. 3A), indicating no lipid peroxidation occurred by any of the tested NMs exposure. Navarro et al. (2012) found that CdSe/ZnS caused oxidative stress in *Arabidopsis thaliana* plants hydroponically cultivated. The different findings are possibly because of the growth media, NMs in soil will undergo aggregation and binding with soil mineral and organic matters, and get less chance to be directly taken up by plants and cause phytotoxicity. In addition, total phenolics, which are class of compounds with the ability of scavenge reactive oxygen species (ROS) and are indicators of the antioxidant stress responses, were unchanged by all types of NMs (Fig. 3B). The unchanged MDA level and total phenolics suggest that the plants had no stress response upon exposure to all types of NMs.

### 3.3. The uptake and translocation of Si, Ti, Zn and Mo in cucumber tissues

Results showed that Si, Ti, Zn in plants root exposed to different types of NMs were not significantly higher than control (Fig. 4), indicating no uptake of NMs or the released ions were taken up by cucumber plants. The results also revealed that the solubility of  $\text{SiO}_2$ ,  $\text{TiO}_2$ , ZnS are extremely low in soil, or the released ions were bound with soil clay minerals lowering their bioavailability. Also, the huge surface area of NPs make it have many active points which

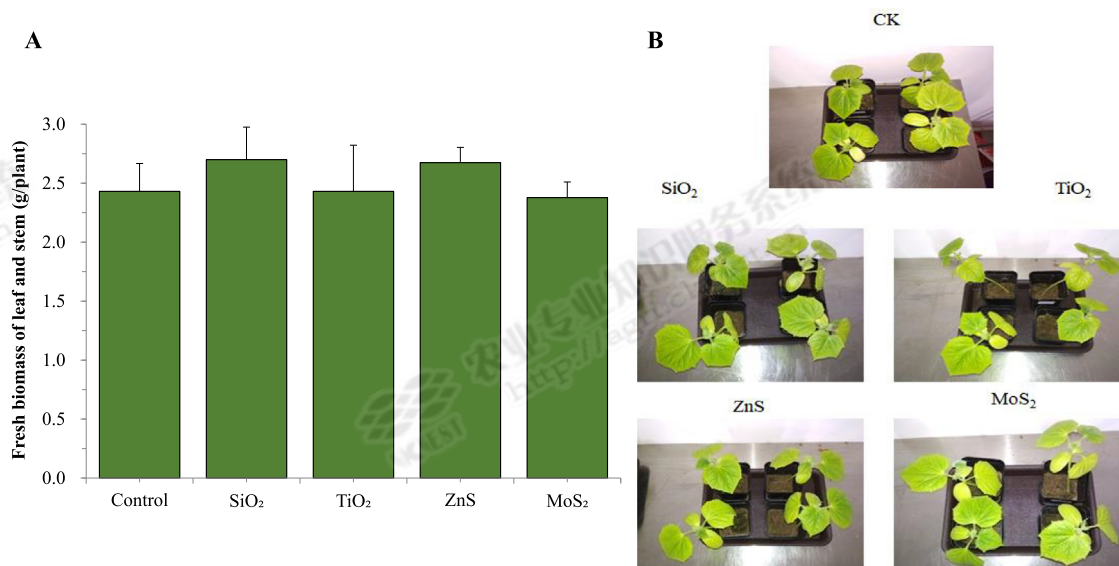


Fig. 2. Cucumber plant above-ground tissues biomass (A) and cucumber images (B).

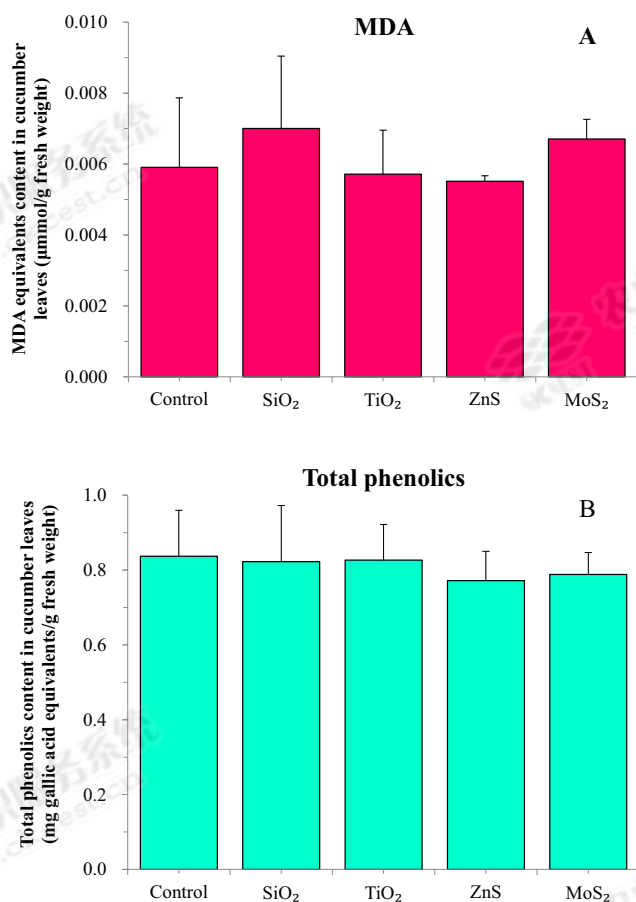


Fig. 3. Impact of Nanomaterials (SiO<sub>2</sub>, TiO<sub>2</sub>, ZnS and MoS<sub>2</sub>) on MDA and total phenolics contents in cucumber plants. All data are average  $\pm$  SD of four replicate.

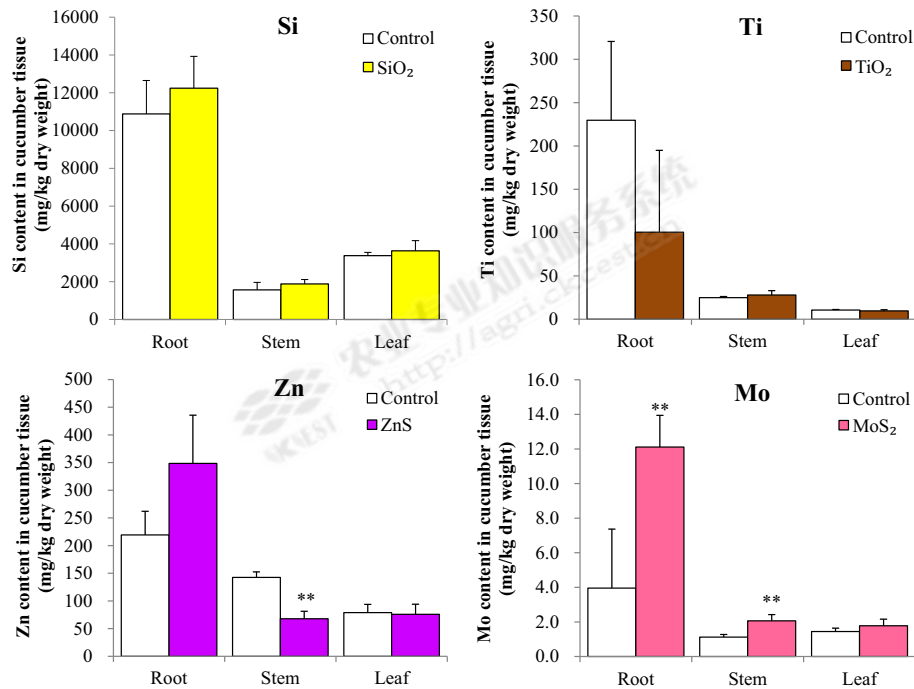
can easily binding with soil minerals or organic matter, and inhibit their uptake and translocate by plants (Song et al., 2013). In above ground tissues (stem and leaf), Si and Ti were also not detected to be higher than that in control, indicating no translocation from root

to upper part tissues (Fig. 4), whereas Zn was observed to be lower than that in control. Aggregation to larger particles which is much bigger than the root pore size is not easy to be taken up by root and translocate to upper part tissues.

The exception is Mo. As shown in Fig. 4, exposure to MoS<sub>2</sub> significantly increased Mo content in root and stem, by 205.6% ( $p < 0.01$ ) and 83.9% ( $p < 0.01$ ), respectively, compared to control. MoS<sub>2</sub> aggregates have large size of more than 1000 nm, which is much bigger than the typical root surface pore size of 20 nm. Therefore, it is almost not possible for plants to uptake MoS<sub>2</sub> aggregates. Thus, we speculate that the Mo in stem is in the form of Mo ion, instead of MoS<sub>2</sub> NMs. Correspondingly, part of the detected Mo in roots should be Mo ions. Molybdenum ions are components of enzymes related to nitrate assimilation and nitrogen fixation, such as nitrate reductase and nitrogenase (Schwarz and Mendel, 2006). This may suggest that MoS<sub>2</sub> NP have the potential to increase nitrate assimilation. Further study is needed to verify this hypothesis.

#### 3.4. Effect of nanomaterials on heavy metals uptake in cucumber plants

Since the collected soils were contaminated by a variety of toxic heavy metals (As, Cd, Cr, Cu, Ni, Al, Zn and Pb), we were curious that whether the amended nanomaterials in soil could decrease the heavy metal uptake in plant tissues due to the adsorption or complexation between NMs and heavy metals. Results showed that heavy metals content (As, Cd, Cr, Cu, Ni, Al, Zn and Pb) in cucumber root exposed to SiO<sub>2</sub> NMs were not significantly changed compared to control (Table 2). This indicates that mesoporous SiO<sub>2</sub> NMs had no impact on heavy metal bioaccumulation in cucumber plant, even previous reports showed mesoporous SiO<sub>2</sub> had good performance in removing heavy metals from wastewater (Sayari et al., 2005; Heidari et al., 2009; Li et al., 2011). While exposure to TiO<sub>2</sub> NMs resulted in a significant reduction uptake of As and Al, by 34.8% and 47.2%, respectively, in cucumber root (Table 2). In addition, exposure to ZnS NMs only significantly ( $p < 0.05$ ) decreased Cr content in cucumber root by 43.3% compared to control (Table 2). Different from other types of NMs, MoS<sub>2</sub> NMs significantly ( $p < 0.05$ ) decreased the content of almost all heavy metals (As, Cd, Cr, Cu, Ni, Al, Ti and Pb) in cucumber roots by approximately



**Fig. 4.** Bioaccumulation of Si, Ti, Zn and Mo in root, stem and leaf of cucumber plants exposed to 100 mg/kg SiO<sub>2</sub>, TiO<sub>2</sub>, ZnS and MoS<sub>2</sub> NMs for 4 weeks. All data are average  $\pm$  SD of four replicate. \*\* represent statistical differences at  $p \leq 0.01$  compared with control, respectively.

**Table 2**

Heavy metal contents in cucumber tissues (mg/kg dry weight). \* and \*\* represent statistical differences at  $p \leq 0.05$  and  $p \leq 0.01$  compared with control, respectively.

|                  | Zn                | Al                | Ti                 | Cu                 | Ni                 | Cr                 | Pb                | As                 | Cd                  |
|------------------|-------------------|-------------------|--------------------|--------------------|--------------------|--------------------|-------------------|--------------------|---------------------|
| Control          | 78.8 $\pm$ 15.0   | 67.0 $\pm$ 11.4   | 10.6 $\pm$ 0.690   | 2.44 $\pm$ 0.231   | 1.88 $\pm$ 0.075   | 0.766 $\pm$ 0.094  | 0.672 $\pm$ 0.148 | 0.380 $\pm$ 0.046  | 0.118 $\pm$ 0.006   |
| SiO <sub>2</sub> | 49.0 $\pm$ 8.35*  | 112 $\pm$ 43.4    | 11.5 $\pm$ 1.03    | 2.35 $\pm$ 0.369   | 2.22 $\pm$ 0.406   | 2.51 $\pm$ 1.16    | 0.682 $\pm$ 0.154 | 0.383 $\pm$ 0.041  | 0.139 $\pm$ 0.039   |
| TiO <sub>2</sub> | 65.1 $\pm$ 9.50   | 81.8 $\pm$ 11.4   | 9.65 $\pm$ 1.25    | 1.79 $\pm$ 0.032** | 1.43 $\pm$ 0.051** | 0.692 $\pm$ 0.109  | 0.647 $\pm$ 0.155 | 0.334 $\pm$ 0.049  | 0.093 $\pm$ 0.011** |
| ZnS              | 75.8 $\pm$ 18.3   | 40.9 $\pm$ 6.33*  | 8.28 $\pm$ 0.650** | 1.73 $\pm$ 0.347*  | 1.52 $\pm$ 0.092** | 0.456 $\pm$ 0.122* | 0.764 $\pm$ 0.139 | 0.297 $\pm$ 0.036* | 0.114 $\pm$ 0.017   |
| MoS <sub>2</sub> | 74.2 $\pm$ 15.2   | 27.7 $\pm$ 11.7** | 8.58 $\pm$ 0.794** | 1.67 $\pm$ 0.026** | 1.69 $\pm$ 0.130   | 3.99 $\pm$ 0.685** | 0.501 $\pm$ 0.094 | 0.299 $\pm$ 0.004* | 0.093 $\pm$ 0.008** |
| Control          | 143 $\pm$ 9.87    | 66.1 $\pm$ 17.9   | 24.9 $\pm$ 1.38    | 13.0 $\pm$ 1.20    | 2.62 $\pm$ 0.268   | 4.26 $\pm$ 0.964   | 2.09 $\pm$ 0.152  | 4.93 $\pm$ 0.869   | 0.677 $\pm$ 0.135   |
| SiO <sub>2</sub> | 58.3 $\pm$ 1.52** | 92.1 $\pm$ 17.3   | 29.8 $\pm$ 2.80*   | 9.70 $\pm$ 0.990** | 2.60 $\pm$ 0.105   | 3.98 $\pm$ 0.453   | 1.87 $\pm$ 0.236  | 2.65 $\pm$ 0.230** | 0.413 $\pm$ 0.033** |
| TiO <sub>2</sub> | 55.7 $\pm$ 13.6** | 93.7 $\pm$ 10.5   | 28.0 $\pm$ 4.97    | 10.1 $\pm$ 1.35*   | 2.48 $\pm$ 0.206   | 3.52 $\pm$ 0.741   | 3.39 $\pm$ 0.754* | 2.43 $\pm$ 0.186** | 0.356 $\pm$ 0.038** |
| ZnS              | 67.9 $\pm$ 13.5** | 60.0 $\pm$ 7.98   | 29.1 $\pm$ 2.17*   | 9.64 $\pm$ 0.948*  | 2.79 $\pm$ 0.488   | 5.28 $\pm$ 1.20    | 1.91 $\pm$ 0.369  | 2.35 $\pm$ 0.354** | 0.411 $\pm$ 0.036** |
| MoS <sub>2</sub> | 50.7 $\pm$ 13.9** | 46.8 $\pm$ 10.7   | 26.9 $\pm$ 4.49    | 12.4 $\pm$ 1.68    | 2.45 $\pm$ 0.207   | 4.15 $\pm$ 0.398   | 2.63 $\pm$ 0.841  | 2.43 $\pm$ 0.350** | 0.358 $\pm$ 0.047** |
| Control          | 219 $\pm$ 42.7    | 7720 $\pm$ 1511   | 230 $\pm$ 91.0     | 30.5 $\pm$ 6.96    | 7.62 $\pm$ 1.78    | 17.5 $\pm$ 3.66    | 201 $\pm$ 63.1    | 22.4 $\pm$ 1.85    | 3.89 $\pm$ 0.565    |
| SiO <sub>2</sub> | 191 $\pm$ 19.1    | 8635 $\pm$ 1273   | 169 $\pm$ 44.0     | 24.7 $\pm$ 2.80    | 5.78 $\pm$ 1.41    | 12.2 $\pm$ 3.07    | 169 $\pm$ 41.4    | 23.6 $\pm$ 5.31    | 4.28 $\pm$ 0.405    |
| TiO <sub>2</sub> | 514 $\pm$ 140*    | 4080 $\pm$ 838*   | 100 $\pm$ 94.5     | 25.0 $\pm$ 4.06    | 6.19 $\pm$ 1.28    | 24.8 $\pm$ 4.04    | 104 $\pm$ 67.9    | 14.6 $\pm$ 3.94*   | 3.20 $\pm$ 0.489    |
| ZnS              | 348 $\pm$ 87.3    | 5894 $\pm$ 600    | 174 $\pm$ 28.0     | 29.4 $\pm$ 5.51    | 5.69 $\pm$ 1.04    | 9.93 $\pm$ 2.38*   | 160 $\pm$ 49.0    | 20.4 $\pm$ 5.92    | 3.39 $\pm$ 0.762    |
| MoS <sub>2</sub> | 203 $\pm$ 36.2    | 3861 $\pm$ 538**  | 90.6 $\pm$ 16.7*   | 19.4 $\pm$ 2.83*   | 3.45 $\pm$ 0.470** | 9.50 $\pm$ 1.98*   | 86.2 $\pm$ 12.4*  | 13.2 $\pm$ 0.802** | 2.41 $\pm$ 0.310**  |

36.4–60.6% relative to control (Table 2). This indicates that MoS<sub>2</sub> NMs have stronger complex or absorption affinity with heavy metal ions compared with other NMs. Wang and Mi (2017) contribute the strong adsorption ability of MoS<sub>2</sub> with heavy metals to the strong Lewis acid and base soft-soft interactions, due to the abundance of exposed sulfur atoms on its surface of MoS<sub>2</sub> NMs.

In stems, exposure to SiO<sub>2</sub> NMs significantly ( $p < 0.01$ ) reduced the content of As (46.2%), Cd (39.0%), Cu (25.4%) and Zn (59.2%) (Table 2). Similarly, exposure to TiO<sub>2</sub> NMs resulted in a significant decrease of the content of As (50.7%,  $p < 0.01$ ), Cd (47.4%,  $p < 0.01$ ), Cu (22.3%,  $p < 0.05$ ) and Zn (61.0%,  $p < 0.01$ ) in stem as well. In addition to SiO<sub>2</sub> and TiO<sub>2</sub> NMs, ZnS also significantly decreased the level of As (52.3%,  $p < 0.01$ ), Cd (39.3%,  $p < 0.01$ ), Cu (25.8%,  $p < 0.05$ ) and Zn (52.5%,  $p < 0.01$ ) in stem. Exposure to MoS<sub>2</sub> NMs significantly decreased the content of As (50.7%,  $p < 0.01$ ), Cd (47.1%,  $p < 0.01$ ) and Zn (64.5%,  $p < 0.01$ ) in stem, with no impact on Cu uptake. In summary, four types of NMs generally had similar pattern on reducing heavy metals bioaccumulation in stem.

In case of leaves, exposure to SiO<sub>2</sub> NMs did not impact the uptake of heavy metals, except Zn, with a 37.8% content decrease ( $p < 0.05$ ) compared to control (Table 2). While TiO<sub>2</sub> NMs resulted in a significant decrease of Cd (21.2%,  $p \leq 0.01$ ), Cu (26.6%,  $p \leq 0.01$ ) and Ni (23.9%,  $p \leq 0.01$ ) content in cucumber leaves compared to control. ZnS NMs significantly reduced content of six heavy metals in cucumber leaves, including As (21.8%,  $p \leq 0.05$ ), Cr (40.5%,  $p \leq 0.05$ ), Cu (29.1%,  $p \leq 0.05$ ), Ni (19.1%,  $p \leq 0.01$ ), Al (39.0%,  $p \leq 0.05$ ) and Ti (21.9%,  $p \leq 0.01$ ) compared to control. MoS<sub>2</sub> at 100 mg/kg decreased accumulation of a number of heavy metals in cucumber leaves, by 21.3% for As ( $p \leq 0.05$ ), 21.2% for Cd ( $p \leq 0.01$ ), 31.6% for Cu ( $p \leq 0.01$ ), 58.7% for Al ( $p \leq 0.01$ ), compared to control without NMs. In contrast, Cr accumulation in cucumber leaves was significantly increased (421%,  $p \leq 0.01$ ) by MoS<sub>2</sub> NMs. Combining that Cr in root was decreased by MoS<sub>2</sub> NMs, the results suggest that MoS<sub>2</sub> NMs enhanced the translocation of Cr from below ground tissues (root) to aboveground tissues (leaf). The underlying mechanism is still unknown.

Combining the data of all tissues, two metal sulfides (ZnS and MoS<sub>2</sub>) had better performance in reducing the heavy metal uptake in cucumber plants compared to metal(loid) oxides NMs (SiO<sub>2</sub> and TiO<sub>2</sub>). Moreover, MoS<sub>2</sub> NMs had more significant adsorption or binding capacity compared with ZnS NMs. The mechanism of the interactions between MoS<sub>2</sub> and adsorbed metal ions has been summarized by Wang and Mi (2017). They assume that the formation of metal-sulfur bonding could be a primary adsorption mechanism. In addition, the electrostatic attraction also contributes to the interaction between ions and MoS<sub>2</sub> (Zhi et al., 2016). Taken together, chemical complexation and electrostatic attraction might be the main mechanism for NMs to complex with heavy metal ions.

### 3.5. Effect of nanomaterials on mineral nutrients uptake in cucumber plants

The adsorption of heavy metals with NMs hindered their uptake and translocation in plant, which is beneficial for crop plant due to the reducing risks to transfer to food chain. However, NMs with high surface area and binding sites, possibly complex with cations like K<sup>+</sup>, Ca<sup>2+</sup> and Fe<sup>3+</sup> as well, and thus lower down the macro- or micro-nutrients uptake by plant, which will be negative impacts for plant growth, given the fact that macro and micro nutrients play an important role in plant metabolism and function. Root is the organ for water and mineral nutrients uptake and leaf is the organ in which important metabolic processes occur, like photosynthesis and respiration. Herein, the excess or deficiency of micronutrient accumulation induced by nanomaterials in root or leaf might perturb plant metabolism and growth.

Concentrations of macro and micro nutrients including Si, Ca, Na, Mg, Fe, K, Mn and Mo found in cucumber tissues grown in soil amended with different types of NMs are shown in Table 3. Exposure to SiO<sub>2</sub> and ZnS generally had no impact on nutrient elements uptake in root compared to control, except that ZnS NMs decreased Mg content by 37.7% ( $p < 0.05$ ). The weak absorbing capacity of SiO<sub>2</sub> and ZnS with mineral nutrient may attribute to their positive surface charge (Table 1). In comparisons, TiO<sub>2</sub> and MoS<sub>2</sub> NMs disturbed the accumulation of a number of nutrients including Si, Mg, Fe, K and Mn in cucumber root. As shown in Table 3, exposure to TiO<sub>2</sub> and MoS<sub>2</sub> NMs significantly ( $p < 0.05$ ) reduced Si by 48.6% and 50.1%, respectively; Mg by 68.4% and 47.8%, respectively; Fe by 45.1% and 48.8%, respectively; K by 59.4% and 49.3%, respectively; Mn by 47.5% and 40.1%, respectively. This indicates that NMs are not only binding with heavy metals, but also with macro and micro nutrients. It is interesting to note that TiO<sub>2</sub> and MoS<sub>2</sub>, which have negative surface charge, induced much more distribution of macro and micro nutrient uptake, compared to two positive charged NMs

(SiO<sub>2</sub> and ZnS). Therefore, surface charge is key factor impacting the interaction between NMs and mineral nutrients. Peralta-Videa et al. (2014) also observed that CeO<sub>2</sub> and ZnO NPs interfered with Mg and K accumulation in soybean root. They proposed the possible underlying mechanism is that NPs stuck on the soybean root surface, blocking the channels and reducing the absorption of Mg. The disturbed nutrient elements (Si, Mg, Fe, K, Mn) act important role in structural integrity, energy storage and redox reaction (Lazar et al., 2003). For example, Mg is a part of the ring structure of the chlorophyll molecule, and the deficiency of Mg will induce chlorosis (Lazar et al., 2003). Mn ions activate several enzymes, such as decarboxylases and dehydrogenases involved in TCA cycle (Peralta-Videa et al., 2014). Therefore, although unchanged at harvest, the biomass might decrease during longer exposure due to the deficiency of these nutrients.

In the above-ground tissues, the most noticeable changes were significantly decreased potassium accumulation in leaves by 8%–17% ( $p < 0.05$ ) when exposed to SiO<sub>2</sub>, TiO<sub>2</sub> and MoS<sub>2</sub>, compared to control (Table 3). Potassium is usually accumulated at high concentrations in plant cells, and K plays an important role in regulation of the osmotic potential of plant cells (Hedrich and Kudla, 2006). It also activates many enzymes involved in respiration and photosynthesis. Therefore, the disturbed K accumulation might negatively impact plant physiological function. As mentioned before, chlorophyll content was negatively impacted by ZnS and MoS<sub>2</sub>, probably because nutrients elements involved in photosynthetic process, such as potassium, were negatively impacted. In addition, exposure to TiO<sub>2</sub>, ZnS and MoS<sub>2</sub> resulted in the significant decrease of leaf Na accumulation by 31.5%–56.8% ( $p < 0.05$ ) compared to control (Table 3). Peralta-Videa et al. (2014) also found that CeO<sub>2</sub> NPs reduced Na accumulation in soybean leaves. Sodium stimulates growth through enhanced cell expansion, and it can partly substitute for potassium as an osmotically active solute (Lazar et al., 2003). Thus, the interfered sodium and potassium uptake in leaf may negatively impact some metabolism process in leaves, like carbon fixation in photosynthesis.

Interestingly, exposure to SiO<sub>2</sub> and TiO<sub>2</sub> NMs significantly ( $p < 0.05$ ) increased Fe content by 58.2% and 40.6%, respectively, in cucumber leaves. Iron plays an important role as a component of enzymes involved in the electrons transfer, such as cytochromes (Lazar et al., 2003). The increased Fe accumulation in leaves is beneficial for photosynthesis process. Another interesting finding is that a significant increase of Si (23%,  $p < 0.01$ ) in leaf was observed in presence of MoS<sub>2</sub> NMs (Table 3). It is known that Si plays an important role in maintain structural integrity and plants deficient in Si are more susceptible to fungal infection. The underlying mechanism is that silicon forms complexes with polyphenols and

**Table 3**  
Mineral nutrients content in cucumber tissues (mg/kg dry weight). \* and \*\* represent statistical differences at  $p \leq 0.05$  and  $p \leq 0.01$  compared with control, respectively.

|                  | Ca           | Mg           | K             | Si           | Na            | Fe           | Mn          | Mo             |
|------------------|--------------|--------------|---------------|--------------|---------------|--------------|-------------|----------------|
| Control          | 28467 ± 3609 | 8085 ± 1304  | 7757 ± 98.9   | 3380 ± 171   | 111 ± 15.0    | 67.7 ± 16.4  | 49.1 ± 9.85 | 1.44 ± 0.201   |
| SiO <sub>2</sub> | 30203 ± 236  | 8877 ± 750   | 7061 ± 340**  | 3634 ± 536   | 107 ± 19.3    | 107 ± 21.5*  | 45.9 ± 2.14 | 1.19 ± 0.228   |
| TiO <sub>2</sub> | 26361 ± 1915 | 7933 ± 663   | 6879 ± 694*   | 3347 ± 292   | 76.5 ± 14.0*  | 95.2 ± 6.57* | 45.3 ± 3.37 | 1.21 ± 0.206   |
| ZnS              | 26629 ± 2240 | 7606 ± 480   | 7232 ± 910    | 4038 ± 373*  | 69.1 ± 21.7*  | 74.0 ± 19.0  | 44.3 ± 4.72 | 0.64 ± 0.069** |
| MoS <sub>2</sub> | 29387 ± 3062 | 7629 ± 803   | 6437 ± 916*   | 4171 ± 119** | 48.4 ± 3.38** | 71.0 ± 14.3  | 45.9 ± 6.93 | 1.78 ± 0.385   |
| Control          | 19886 ± 1559 | 8455 ± 1126  | 13857 ± 1188  | 1566 ± 398   | 1374 ± 267    | 119 ± 25.4   | 20.3 ± 3.59 | 1.12 ± 0.155   |
| SiO <sub>2</sub> | 23853 ± 3390 | 9771 ± 1048  | 14135 ± 1459  | 1879 ± 233   | 1811 ± 173    | 110 ± 18.2   | 23.8 ± 2.09 | 1.14 ± 0.104   |
| TiO <sub>2</sub> | 21189 ± 1294 | 8425 ± 496   | 14199 ± 1669  | 1904 ± 249   | 1756 ± 254    | 117 ± 9.73   | 23.3 ± 2.66 | 1.03 ± 0.109   |
| ZnS              | 22062 ± 1856 | 8738 ± 404   | 17375 ± 1606* | 1816 ± 86    | 1768 ± 129*   | 94.7 ± 8.09  | 21.9 ± 1.85 | 0.77 ± 0.059** |
| MoS <sub>2</sub> | 21476 ± 2352 | 8312 ± 1678  | 15303 ± 3000  | 1560 ± 139   | 1923 ± 157*   | 96.5 ± 20.2  | 20.2 ± 1.75 | 2.06 ± 0.360** |
| Control          | 13707 ± 1460 | 3422 ± 553   | 2685 ± 760    | 10880 ± 1773 | 699 ± 163     | 7102 ± 1129  | 339 ± 68.4  | 3.96 ± 3.41    |
| SiO <sub>2</sub> | 14958 ± 1241 | 2742 ± 162   | 1822 ± 305    | 12245 ± 1683 | 616 ± 124     | 7714 ± 1187  | 379 ± 77.6  | 1.13 ± 0.493   |
| TiO <sub>2</sub> | 15257 ± 2796 | 1080 ± 347** | 1091 ± 462*   | 5589 ± 1182* | 664 ± 139     | 3897 ± 1144* | 178 ± 68.5* | 11.0 ± 8.42    |
| ZnS              | 13694 ± 602  | 2131 ± 427*  | 1617 ± 493    | 8340 ± 710   | 562 ± 71.1    | 5355 ± 229   | 278 ± 7.25  | 3.70 ± 3.23    |
| MoS <sub>2</sub> | 14071 ± 972  | 1781 ± 274** | 1362 ± 233*   | 5431 ± 735** | 581 ± 112     | 3638 ± 389** | 203 ± 41.0* | 12.1 ± 1.83**  |

serve as an alternative to lignin in the reinforcement of cell wall (Lazar et al., 2003). Therefore, these results suggest that MoS<sub>2</sub> might help plant resist pests or fungus through the enhancement of Si uptake in cucumber leaves.

#### 4. Conclusions

In summary, the results of this study indicate that four types of nanomaterials dosing as low as 100 mg/kg, have the potential to reduce toxic heavy metal accumulation in cucumber plants. In the experimental conditions of this study, MoS<sub>2</sub> NMs impacted the accumulation of most heavy metals. Thus, the tested NMs especially MoS<sub>2</sub> NMs, can be applied as immobilization amendment in contaminated soils to reduce the risk of the transfer of toxic heavy metals to food chain. However, although the biomass was unchanged by NMs, the photosynthetic pigments contents were decreased by metal sulfides NMs (MoS<sub>2</sub> and ZnS). Additionally, NMs also inhibited the uptake of macro and micro nutrients in cucumber plants. The negative impact on beneficial nutrients involved in photosynthesis, such as Fe and Mg, may impact physiological function. The uptake and translocation of NMs or their released ions by plant tissues were not found, except MoS<sub>2</sub>/Mo. In presence of MoS<sub>2</sub>, the content of Si in cucumber leaves increased, which may enhance the resistance to attackers. This study provides a perspective on how emerging nanomaterials impact the heavy metals uptake and also physiological impact on plants. Both positive (reduce heavy metal accumulation) and negative (decrease nutrient element accumulation) impact should be taken into consideration when NMs were applied in environmental remediation. Cucumber plants, which is sensitive to heavy metal contaminations, can be applied with NMs, especially MoS<sub>2</sub>, to enhance their tolerance and resistance to heavy metals.

#### Acknowledgements

This work was funded by the National Key Research and Development Program of China under 2018YFD0201000 and National Natural Science Foundation of China under 31771726. Any opinions, finding, and conclusions or recommendations expressed in this material are those of authors and do not necessarily reflect the views of National Science Foundation of China.

#### References

- Bolan, N., Kunhikrishnan, A., Thangarajan, R., Kumpiene, J., Park, J., Makino, T., Kirkham, M.B., Scheckel, K., 2014. Remediation of heavy metal(loid)s contaminated soils – to mobilize or to immobilize? *J. Hazard. Mater.* 266, 141–166.
- Cachada, A., Rocha-Santos, T., Duarte, A.C., 2018. Chapter 1 - soil and pollution: an introduction to the main issues. In: Duarte, A.C., Cachada, A., Rocha-Santos, T. (Eds.), *Soil Pollution*. Academic Press, pp. 1–28.
- Chen, X., Lam, K.F., Yeung, K.L., 2011. Selective removal of chromium from different aqueous systems using magnetic MCM-41 nanosorbents. *Chem. Eng. J.* 172 (2–3), 728–734.
- Chen, X., Lam, K.F., Zhang, Q., Pan, B., Arruebo, M., Yeung, K., 2009. Synthesis of highly selective magnetic mesoporous adsorbent. *J. Phys. Chem. C* 113 (22), 9804–9813.
- Fan, X., Wang, C., Wang, P., Hu, B., Wang, X., 2018. TiO<sub>2</sub> nanoparticles in sediments: effect on the bioavailability of heavy metals in the freshwater bivalve *Corbicula fluminea*. *J. Hazard. Mater.* 342, 41–50.
- González, V., Díez-Ortiz, M., Simón, M., Van Gestel, C.A.M., 2013. Assessing the impact of organic and inorganic amendments on the toxicity and bioavailability of a metal-contaminated soil to the earthworm *Eisenia andrei*. *Environ. Sci. Pollut. Res. Int.* 20 (11), 8162–8171.
- Gorsuch, J.W., Lower, W.R., Lewis, M.A., Wang, W., 1991. *Plants for Toxicity Assessment*, ASTM STP 11152. ASTM, Philadelphia.
- Hedrich, R., Kudla, J., 2006. Calcium signaling networks channel plant K<sup>+</sup> uptake. *Cell* 125 (7), 1221–1223.
- Heidari, A., Younesi, H., Mehraban, Z., 2009. Removal of Ni(II), Cd(II), and Pb(II) from a ternary aqueous solution by amino functionalized mesoporous and nano mesoporous silica. *Chem. Eng. J.* 153 (1–3), 70–79.
- Jambunathan, N., 2010. Determination and detection of reactive oxygen species (ROS), lipid peroxidation, and electrolyte leakage in plants. In: Sunkar, R. (Ed.), *Plant Stress Tolerance: Methods and Protocols*. Humana Press, Totowa, NJ, pp. 291–297.
- Konate, A., He, X., Rui, Y.K., Zhang, Z.Y., 2017. Magnetite (Fe<sub>3</sub>O<sub>4</sub>) nanoparticles alleviate growth inhibition and oxidative stress caused by heavy metals in young seedlings of cucumber (*Cucumis Sativus L.*). *ITM Web of Conferences* 12, 03034.
- Lazar, T., Taiz, L., Zeiger, E., 2003. *Plant Physiology*, third ed., vol. 91, pp. 750–751 NonEn.
- Lehman, S.E., Larsen, S.C., 2014. Zeolite and mesoporous silica nanomaterials: greener syntheses, environmental applications and biological toxicity. *Environ. Sci. Nano* 1 (3), 200–213.
- Li, G., Zhao, Z., Liu, J., Jiang, G., 2011. Effective heavy metal removal from aqueous systems by thiol functionalized magnetic mesoporous silica. *J. Hazard. Mater.* 192, 277–283.
- Lin, Y.S., Haynes, C.L., 2009. Synthesis and characterization of biocompatible and size-tunable multifunctional porous silica nanoparticles. *Chem. Mater.* 21 (17), 3979–3986.
- Navarro, D.A., Bisson, M.A., Aga, D.S., 2012. Investigating uptake of water-dispersible CdSe/ZnS quantum dot nanoparticles by *Arabidopsis thaliana* plants. *J. Hazard. Mater.* 211–212, 427–435.
- Pala, I.R., Brock, S.L., 2012. ZnS nanoparticle gels for remediation of Pb<sup>2+</sup> and Hg<sup>2+</sup> polluted water. *ACS Appl. Mater. Interfaces* 4 (4), 2160–2167.
- Peralta-Videa, J.R., Hernandez-Viezcas, J.A., Zhao, L., Diaz, B.C., Ge, Y., Priester, J.H., Holden, P.A., Gardea-Torresdey, J.L., 2014. Cerium dioxide and zinc oxide nanoparticles alter the nutritional value of soil cultivated soybean plants. *Plant Physiol. Biochem.* 80, 128–135.
- Praveen, A., Khan, E., Ngiime, D.S., Perwez, M., Sardar, M., Gupta, M., 2018. Iron oxide nanoparticles as nano-adsorbents: a possible way to reduce arsenic phytotoxicity in Indian mustard plant (*Brassica juncea L.*). *J. Plant Growth Regul.* 37 (2), 612–624.
- Sayari, A., Hamoudi, S., Yang, Y., 2005. Applications of pore-expanded mesoporous silica. 1. Removal of heavy metal cations and organic pollutants from wastewater. *Chem. Mater.* 17 (1), 212–216.
- Schwarz, G., Mendel, R.R., 2006. Molybdenum cofactor biosynthesis and molybdenum enzymes. *Annu. Rev. Plant Biol.* 57 (1), 623–647.
- Šesták, Z., Catský, J., Jarvis, P.G., 1971. Plant photosynthetic production. In: Junk NV, W. (Ed.), *Manual of Methods*. The Hague, Netherlands, p. 818.
- Singleton, V.L., Rossi, J.A., 1964. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am. J. Enol. Vitic.* 16 (3), 144–158.
- Song, U., Shin, M., Lee, G., Roh, J., Kim, Y., Lee, E.J., 2013. Functional analysis of TiO<sub>2</sub> nanoparticle toxicity in three plant species. *Biol. Trace Elem. Res.* 155 (1), 93–103.
- Stark, W.J., Stoessel, P.R., Wohlleben, W., Hafner, A., 2015. Industrial applications of nanoparticles. *Chem. Soc. Rev.* 44 (16), 5793–5805.
- Tripathi, D.K., Singh, V.P., Prasad, S.M., Chauhan, D.K., Dubey, N.K., 2015. Silicon nanoparticles (SiNp) alleviate chromium (VI) phytotoxicity in *Pisum sativum* (L.) seedlings. *Plant Physiol. Biochem.* 96, 189–198.
- Wang, Z., Mi, B., 2017. Environmental applications of 2D molybdenum disulfide (MoS<sub>2</sub>) nanosheets. *Environ. Sci. Technol.* 51 (15), 8229–8244.
- Yue, L., Chen, F., Yu, K., Xiao, Z., Yu, X., Wang, Z., Xing, B., 2019. Early development of apoplastic barriers and molecular mechanisms in juvenile maize roots in response to La<sub>2</sub>O<sub>3</sub> nanoparticles. *Sci. Total Environ.* 653, 675–683.
- Zhao, L., Zhang, H., White, J.C., Chen, X., Li, H., Qu, X., Ji, R., 2019. Metabolomics reveal that engineered nanomaterial exposure in soil alters both soil rhizosphere metabolite profiles and maize metabolic pathway. *Environ. Sci.: Nano* 6, 1716–1727. In this issue.
- Zhi, L., Zuo, W., Chen, F., Wang, B.D., 2016. 3D MoS<sub>2</sub> composition aerogels as chemosensors and adsorbents for colorimetric detection and high-capacity adsorption of Hg<sup>2+</sup>. *ACS Sustain. Chem. Eng.* 4 (6), 3398–3408.
- Zhu, Y., Yu, H., Wang, J., Fang, W., Yuan, J., Yang, Z., 2007. Heavy metal accumulations of 24 asparagus bean cultivars grown in soil contaminated with Cd alone and with multiple metals (Cd, Pb, and Zn). *J. Agri. Food Chem.* 55 (3), 1045–1052.